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Carboxylate and Phosphonate Modified Polyethyleneimine: Towards the Design of Actinide Decorporation Agents.

Florian Lahrouch^{†/}, Bruno Siberchicot[‡], Jeanne Fèvre[†], Laurane Leost[†], Jean Aupiais[‡], Pier Lorenzo Solari[§], Christophe Den Auwer^{†*} and Christophe Di Giorgio^{†**}

[†] Université Côte d'Azur, CNRS, Institut de Chimie de Nice, UMR 7272, 06108 Nice, France

[‡] CEA, DAM, DIF, F-91297 Arpajon cedex, France

[§] Synchrotron SOLEIL, L'Orme des Merisiers, Saint-Aubin, BP 48, F-91192 Gif-sur-Yvette, France

/ present address : Sorbonne Université, IMPMC UMR 7590, Paris, France

* christophe.denauwer@univ-cotedazur.fr

* christophe.di-giorgio@univ-cotedazur.fr

ABSTRACT

Plutonium (Pu) is an anthropogenic element involved in the nuclear industry cycle. Located at the bottom of the periodic table within the actinide family, it is a chemical toxic but also a radiological toxic, regardless of isotopy. After nearly 80 years of plutonium industrialization, it has become clear that inhalation and wounds represent the two main ways a person may become contaminated after an accident. In order to reduce the deleterious health effects of Pu, it is crucial to limit chronic exposure by removing it or preventing its incorporation into the body. Diethylene triamine penta acetic acid (DTPA) has emerged as the gold standard for plutonium decorporation, although it suffers from very short retention time in serum. Other molecules like the hydroxypyridonate family with high chemical affinity have also been considered. We have been considering alternative polymeric chelates and, in particular, polyethyleneimine (PEI) analogues of DTPA (carbonate or phosphonate version), which may present a real breakthrough in plutonium decorporation not only because of their higher loading capacity, but also because of their indirect vectorization properties correlated with a specific biodistribution into the lungs, bone, kidney or liver.

In the first part of this *forum* article, new data on the structural characterization of the complexation of Pu(IV) with polyethyleneimine methylphosphonate (PEI-MP) were obtained using the combination of EXAFS spectroscopy (Extended X-Ray Absorption Fine Structure) and *ab initio* Molecular Dynamics calculations (AIMD). The use of thorium (Th) as a Pu chemical

surrogate is also discussed because its unique oxidation state is +IV in solution. In the second part of the paper, we put this new set of data on PEI-MP-Pu in perspective with the use of the PEI platform to complex Th(IV) and Pu(IV). Uptake curves of Th(IV) with PEI-MC, PEI-MP and DTPA are compared and *ab initio* Molecular Dynamics data are discussed.

INTRODUCTION

Since the end of the Second World War, the development of civil and military nuclear industry in countries of the northern hemisphere has brought some new issues in terms of environmental impact, legacy and long-term management, defense and non-proliferation. The Chernobyl accident in April 1986 and more recently the Fukushima accident in March 2011 have had a major impact in terms of energetic policies in various countries, mostly in the West, because of the decrease in social acceptance of civil nuclear power.¹ Forensics and possible misuse of nuclear materials for terrorism is also being questioned lately. In 2017, the Nobel Peace Prize was given to ICAN (International Campaign to Abolish Nuclear weapons) demonstrating the growing concern about the former Cold War atomic race and non proliferation. Plutonium (Pu) is an issue of both nuclear weapon stockpiles, forensics and energy sources of nuclear fuel. Furthermore, plutonium is probably the element associated with most societal questioning and fear. In addition, atomic tests performed in atmospheric conditions from 1945 (USA) to 1980 (China) have deposited several tons of radionuclides with highly anisotropic repartition throughout the surface of the planet. It is for instance approximately $6.52 \cdot 10^{15}$ Bq (i.e. ≈ 3 tons) of ^{239}Pu that was released in the atmosphere during the second half of the 20th century.²

In environmental conditions, plutonium is mostly present at the +IV oxidation state under soluble and/or colloidal forms (although oxidation states +III and +V must not be excluded in reducing or oxidative conditions, respectively). Given the severe chemical and radiotoxicity of all isotopes of plutonium, its handling, utilization and possible accidental dispersion raises public health issues especially in the case of internal contamination. It was within the framework of the Manhattan Project (1942 – 1946) that the investigation of the health effects associated with radiochemistry and weapons production was initiated. In 1943, the same year as the construction of the full-scale plutonium production reactor (Hanford B reactor, WA, USA), the University of Rochester was commissioned by the Atomic Energy Commission to assess the potentially deleterious effects of atomic energy production on human health. In 1962, Professor Harold C. Hodge created the first Society of Toxicology (SOT).³

After nearly 80 years of plutonium industrialization, inhalation and wounds represent the two main entryways of accidental contamination.⁴ It is important to note that generally speaking plutonium (and thorium) is not naturally excreted from the body because its retention time is longer than human life expectancy. After contamination, the blood system acts as the major distribution vector of plutonium to organs⁵ and the kinetics of this transfer is relatively fast because 1 day after contamination less than 1% of injected Pu is still in the serum.⁶ In this case, the soluble fraction of plutonium as well as the smallest Pu particles are able to pass into the blood

system.⁷ The skeleton and liver are affected by the retention of plutonium although they exhibit differences in their biodistribution.⁶ The soluble phase has a strong affinity with the highly vascularized hematopoietic marrow located in the bones⁸ and for the ferric proteins such as ferritin⁹, largely present in liver. The colloidal forms of plutonium are managed by the reticuloendothelial system also located in the liver and bones.⁵ Furthermore, the continuous irradiation of internal tissues caused by the severe radiotoxicity of plutonium is at the origin of serious tissue damage and can trigger bone and liver cancers or leukaemia.^{10, 11} In the case of inhalation, the insoluble portion as well as the colloids which are not able to translocate to the blood compartment remain trapped into alveolar macrophages which serve as a reservoir for actinide retention.¹² To reduce the deleterious health effects of Pu, it is crucial to remove or prevent incorporation in the body in order to limit the long-term chronic exposure. However, the partial hydrolysis of actinides at physiological pH, their interactions with biologic matter (proteins, metabolites, bone matrix, liver, macrophages, etc.) or the abundance of endogenous cations (Fe(III), Ca(II)) make the specific actinide extraction from the biological media difficult. Additionally, their different chemical properties require the development of specific chelators adapted to each actinide.

First attempts of Pu human decorporation treatments used dietary modification or hormones.¹³ But this proved to be unsuccessful. Painter *et al.* administrated sodium citrate to Pu(IV) contaminated dogs and observed an increase of urinary Pu(IV) excretion.¹⁴ Dimercaptopropanol (BAL), usually used to cure the heavy metal poisoning (lead, mercury or bismuth), was for a time envisioned for plutonium and radium decorporations.¹⁴ Eventually, BAL was revealed to be more effective for polonium chelation.¹⁵ Amino carboxylate acids started to be studied for actinide chelation in the 50's. First, ethylene diamine tetra acetic acid (EDTA) which forms a stable complex with Pu(IV) ($\log\beta = 26.4$)¹⁶ was tested as calcium salts (CaNa₂-EDTA) but its lack of selectivity for Pu(IV) prevented its use.¹⁷ Then the diethylene triamine penta acetic acid (DTPA) emerged as a powerful Pu(IV) chelator. The complexation site formed by 5 oxygen and 3 nitrogen atoms exhibits a high chelating constant for plutonium (IV) ($\log\beta = 36.36$, $I=0.1M$).^{18,19} Studies on mice showed that injections of Ca-DTPA, less than one hour after contamination, permit the urinary excretion of around 60% of the inner Pu(IV).²⁰ Also DTPA modifies the Pu(IV) biodistribution limiting the bone and liver retention.²⁰ Still used today, DTPA injected as Ca(II) or Zn(II) salt form is considered as the most effective chelating agent for promoting Pu(IV) excretion²¹. However, this molecule is severely limited because of its weak selectivity towards Pu and its narrow distribution within the body,²²⁻²⁵ being active mainly in serum^{20,26} and for a short period of time (from a report of Breustedt and coworkers, 400 min after injection, less than 1% of free DTPA remains in the blood compartment²⁷). As already mentioned above, although the retention time of Pu(IV) in serum is less than a few hours, a remedy is needed for the fraction

which does not enter into circulation, i.e. lung and their macrophages, or go beyond the blood system to target bone or liver.⁶

Modern research on new chelating functions with affinity for Pu(IV) and other actinides such as Am(III) or Np(V) lead to the development of catecholate (CAM) and hydroxypyridonate (HOPO) templates inspired by the siderophore family.²⁸ These functional groups attached to spatially suitable molecular backbones to form multidentate chelating agents were largely studied by Raymond and Durbin research groups. Among the developed ligands, the octadentate 3,4,3-LI(1,2-HOPO) emerged as a lead candidate for actinide decorporation therapy forming with Pu(IV) very stable complexes.²⁹ The first *in vivo* studies showed that after the mice were contaminated, the 3,4,3-LI(1,2-HOPO) ligand was able to modify the biodistribution of Pu(IV), Am(III) and Np(V) and to drastically reduce bone, liver and soft tissues retention.³⁰ 3,4,3-LI(1,2-HOPO) is also well tolerated in rats at very high doses ($100 \mu\text{mol kg}^{-1} \text{ day}^{-1}$) for 28 days of daily oral administration.³¹ Moreover, an *in vitro* study showed that the 3,4,3-LI(1,2-HOPO) ligand does not disturb the activity of human liver microsomes (pool composed of phenacetin, bupropion, diclofenac, mephentyoin, bufuralol and testosterone), it is neither sensitive to the enzymatic oxidation occurring in human plasma nor to the acidic conditions present in the gastro-intestinal tract.³² Moreover, the 3,4,3-LI(1,2-HOPO) also seems able to form stable complexes with a large variety of actinides.³³

So far, the development of these molecular strategies has proven very efficient in optimizing the affinity constants with actinides. But this strategy somehow finds its limitations when the actinide element is already out of the blood system or incorporated into the target organs. An alternative to this molecular strategy would be to enhance organ tropism using vectorization of molecular chelating agents. Some formulations have been seen to promote extra-blood system biodistribution and to facilitate administration. This is the case for the lipophilic form of DTPA³⁴⁻³⁷ or Nano-DTPA®³⁸, which consists of encapsulating DTPA with zinc acetate and other adjuvants. The latter was tested on dogs in order to decorporate Am(III).³⁹ Liposomal forms of DTPA were recently considered using polyethylene glycol PEG to enhance the retention time and the biodistribution of DTPA.⁴⁰⁻⁴²

Our research group is focusing on polymeric chelates and more particularly on polymeric analogues of DTPA, which may represent a real breakthrough in the actinide decorporation field because of their higher loading capacity (larger abundance of chelating sites per mg of polymer that could enhance uptake rates), but mostly because of their indirect vectorization properties correlated with a specific biodistribution into the lungs, bone, kidney or liver.⁴³ This tropism is mainly structural, i.e. polymers are retained in the main organs due to a size effect and/or enhanced binding properties (ionic, hydrogen, van der Waals) and are more prone to activate phagocytosis from macrophages. Our polymeric structural analogues of DTPA are therefore based

on the well-known branched polyethyleneimine (bPEI) molecule either functionalized with methylcarboxylates (true higher homologue) or with methylphosphonates (used as another class of chelating groups). Studies performed with alkylcarboxylated PEI⁴⁴ or succinamic acid PEI⁴⁵ revealed that anionization of PEI resulted into improved biocompatibility and decreased the cytotoxicity of the related modified polymers. Recently, we have reported that the functionalized polyethyleneimine methylcarboxylate (PEI-MC, Figure 1a) is able to complex Pu(IV) and Th(IV) by forming the same complexation site as the DTPA.⁴⁶ PEI-MC is also able to complex U(VI) under physiological model conditions.⁴⁷ As for the polyethyleneimine methylphosphonate (PEI-MP, Figure 1b), it is known to seek out bone macromolecules and was at one time considered for ^{117m}Sn or ^{99m}Tc complexation for bone cancer imagery and scintigraphy.⁴⁸ Biodistribution and biokinetic studies have shown that the PEI-MP is distributed to the bones and liver with a biological half time of less than 30 min in the liver and more than 4 h in the bone.⁴⁹ PEI-MP was also investigated for its capacity to complex U(VI) in pseudo physiological conditions.⁵⁰

This *forum* article is formally divided in two parts. First, we present new results on the structural characterization of the complexation of Pu with polyethyleneimine methylphosphonate (PEI-MP) using the combination of EXAFS spectroscopy (Extended X-Ray Absorption Fine Structure) and *ab initio* molecular dynamics calculations (AIMD). We also discuss the use of thorium (Th) as a plutonium chemical analogue (surrogate) because its unique oxidation state is +IV in solution and also because its specific activity (for ²³²Th, 4×10³ Bq.g⁻¹) makes it much easier to manipulate in the laboratory than Pu, regardless of its isotopy. Th has frequently been reported to mimic the chemistry of plutonium at oxidation state +IV although this must always be taken with caution.⁵¹ One limitation comes from the increase of the nuclear charge that is in brief responsible for radii contraction occurring from the left to the right of the series together with relative stabilization of the 5f orbitals. Consequently, the physical chemical properties of Pu differ slightly from that of Th, even at the same oxidation state +IV. Nevertheless, the physical chemical data for Th and Pu presented here represent a basic chemistry approach necessary for further biological evaluations of these modified polymers towards Pu(IV) contamination. The second part of the paper puts this new set of data on PEI-MP-Pu in perspective with the use of the PEI platform to decorporate actinides. Previous reported characterizations for both PEI-MP and PEI-MC involved in the complexation of M(IV) (M = Th and Pu) are also discussed.

EXPERIMENTAL SECTION

The PEI-MP was synthesized as described in a previous study.⁵⁰

Uptake curves

The experimental conditions for the measurement of the uptake curves of PEI-MP-Th, PEI-MC-Th and DTPA-Th are described in previous reports.^{46, 50}

Each series of Th(IV) uptake data points was fitted to a four-parameter logistic nonlinear regression model⁵² representing a truncated sigmoid:

$$y = a + \frac{(b - a)}{\left[1 + \frac{EC50^n}{x^n}\right]}$$

where y is the expected response at dosage step x , a is the response at step = 0, b is the maximum response at infinite step, $EC50$ is the titration point at 50% response (it also reflects the inflection point of the dose–response curve) and n is the slope at the $EC50$.

For both systems, a was kept at 5% according to the result of the blank found for Th(IV) in the absence of a chelating agent.

Extended X-ray absorption fine structure (EXAFS) of PEI-MP-Pu

PEI-MP-Pu Sample preparation

25 μ L of a solution of plutonium nitrilotriacetate solution ($[Pu] = 5 \times 10^{-4}$ M, pH 4) was added to 225 μ L of a PEI-MP solution (1 mg.mL⁻¹) containing TBS buffer (pH = 1.0, $[TRIS] = 5 \times 10^{-5}$ M, $[NaCl] = 15 \times 10^{-5}$ M, pH=7). The final pH was equal to 6.5 and Pu final concentration equal to 5.5×10^{-5} M. This corresponds to a stoichiometry of the Pu:PEI-MP monomer molar ratio of 0.02 (this large under-stoichiometry of Pu ensures that the limit complex was obtained).

Data recording and processing

EXAFS experiments at the Pu L_{III} edge were carried out on the MARS beamline of the Soleil synchrotron facility. The MARS beamline is dedicated to the investigation of radioactive materials in the hard X-ray range. Because of the low concentration of the samples, EXAFS measurements were performed in fluorescence mode using a 13-element high purity germanium detector (ORTEC) with specifically designed 200 μ L cells (Ets CANAPLE, Grenoble, Fr). The optics of the beamline essentially consist of a water-cooled double-crystal monochromator (FMB Oxford), which is used to select the incident energy of the X-ray beam and for horizontal focalization, and two large water-cooled reflecting mirrors (IRELEC/SESO) that are used for high-energy rejection (harmonic portion) and vertical collimation and focalization. In this case, the monochromator was set with the Si (220) crystals and the mirrors with the Pt strips. Energy calibration was performed at the Y K edge at 17038 eV.

Data processing was carried out using the ATHENA code.⁵³ The e_0 energy was identified at the maximum of the absorption edge data fitting. The signal to noise of the average spectrum (0.016 for 7 scans) is relatively low because of the low concentration of Pu in the EXAFS cells. It is estimated that the final concentration of Pu during beam time measurement was below 5×10^{-5} M because sorption of Pu(IV) onto the vessel between sample preparation and sample measurement always decreases the nominal concentration.

Data fitting

Fourier Transformation (FT) in k^2 was performed between 3 and 10 $\text{\AA}^{-1}$ with Hanning windows using the ARTEMIS code.⁵³ This short range is due to the low concentration of Pu in the sample cell triggering strong noise on the EXAFS data ($\epsilon(k)=0.016$). Only one global amplitude factor S_0^2 and one energy threshold correction factor ΔE_0 were used for all paths. Phases and amplitudes were calculated using the FEFF6 simulation code⁵⁴ included in ARTEMIS and based on the structure of the PEI-MP model monomer complexed to Pu(IV) obtained by the quantum molecular dynamics.

Quantum molecular dynamics

In the framework of DFT-PAW formalism, we used the ABINIT code⁵⁵ for performing AIMD simulations on both Th and Pu-PEIMP models. For each case, we treated an isolated charged complex in a large simulation box in order to simulate the gaseous phase system. We also performed simulation in the presence of 22 water molecules in order to mimic the solvent effects. The simulations are run in the isokinetic ensemble at 298 K until equilibration is achieved, i.e. until the variations in pressure stabilize to an oscillatory pattern, without any long-term trend. The size of the simulation cell (20 Bohr) is chosen in order to obtain a pressure close to zero. Following equilibration, the pressure and energy are calculated by averaging over the next 2000 time steps of 0.48 fs each. All calculations were done at the Γ point, the centre of the Brillouin zone. For carbon, oxygen and hydrogen the pseudo potentials were taken from the ABINIT data base⁵⁶, for thorium and plutonium a PAW data set was fixed and a pseudo-potential built and checked.⁵⁷ The data were generated for the Perdew-Burke-Ernzerhof (PBE) exchange-correlation functional with relativistic corrections to the wave function including spin-orbit coupling.⁵⁸ We have chosen a standard actinide basis set with 7s, 6p, 6d, 5f in the valence band and 6s and 6p as the semicore orbitals. In ABINIT the energy cut off is an important parameter and we investigated the convergence needed for obtaining reliable results. This cut off was fixed at 760 eV.

In plutonium systems, the presence of correlated 5f electrons leads to an improper description of the electronic and the structural properties by density functional theory. To overcome these shortcomings of DFT the DFT+U (Generalized Gradients Approximation GGA+U) approach has proven to effectively correct the deficiencies. Subsequently, except for thorium (no 5f electron in Th(IV)), we introduced Hubbard U and Hund's J parameters for the onsite interaction strength. Although the choice of U and J are debatable we used the classical values obtained for tetravalent plutonium oxide ($U = 4$ eV, $J = 0.2$ eV).⁵⁹

RESULTS ON THE PEI-MP-Pu(IV) SYSTEM.

Structural investigation of the complexation site of Pu(IV) with PEI-MP was performed with EXAFS spectroscopy (at the Pu L_{III} edge) in combination with AIMD simulations. EXAFS spectra

were recorded in pseudo physiological conditions (pH = 6.5, 150 mM NaCl) with a Pu: PEI-MP monomer molar ratio of 0.02 as explained in the experimental section (Figure 2a). The corresponding modulus of the Fourier transform (FT) is shown in Figure 2b. The best fit metrical parameters of the PEI-MP-Pu spectrum are displayed in Table 1 as well as comparison with data reported previously on the parent complex PEI-MP-Th. The first observation is a contraction of the first M-O coordination sphere from M = Th (2.35(1) Å) to M = Pu (2.26(1) Å) ($\Delta = -0.09$ Å) in agreement with the so called actinide contraction of the ionic radii from light to heavy elements of the actinide family. Note that the number of O atoms was fixed arbitrarily to 9 in both cases because the adjustment of absolute coordination numbers would be meaningless in the absence of reliable model compounds (as it is often the case for Pu(IV) and to lesser extent for Th(IV)). In the second coordination sphere, phosphate ligation is observed for both spectra with much a larger bond distance contraction ($\Delta = -0.28$ Å) from Th (3.90(1) Å) to Pu 3.62(12) Å. The great uncertainty associated with the Pu---P distance is due to the lower signal to noise ratio of the spectrum (see experimental section), indicating that data on the second coordination sphere must be discussed with much care. Last, in the fit, the presence of a Pu---Cl contribution is compulsory as was already observed for the fit of PEI-MP-Th.⁵⁰

Ab initio Molecular Dynamics simulations were performed at 298 K by the addition of 6 water molecules for PEI-MP*-M (M = Th, Pu, PEI-MP* = model monomeric unit of PEI-MP terminated with an ethyl and a methyl group), using ABINIT code and will be fully discussed for the comparison between Th and Pu in the next section.⁵⁵ The monomer, presented in Figure 1a was selected to be a representative of the entire PEI-MP polymer which was obtained by the functionalization of the commercial bPEI.⁴⁶ Following the natural distribution of the amines of type I, II and III (approximately equal to 0.33/0.33/0.33 respectively in this commercial 25 kDa branched bPEI⁶⁰), this average model monomer is composed of three phosphonate functional groups and three nitrogen atoms. After 1 ps of equilibration, three water molecules came to complete the first cation coordination shell. A snapshot image of the PEI-MP*-Pu complex is displayed in Figure 3a. Average distances are reported in Table 3 based on radial pair distribution functions $g(r)$. In Figure 4c, the first Pu-O shell is split between 3 O atoms at 2.20 Å (corresponding to the 3 O atoms from phosphonate groups) and 3 O at 2.50 Å (for the 3 water molecules). The average equals 2.35 Å. *ab initio* MD simulations also indicate that 1 N atom from the amino backbone of PEI-MP* is involved in the Pu coordination at 2.65 Å. But the presence of one extra N atom in the same shell is unlikely to be detected in EXAFS given the similarity of atomic numbers between O and N. This contribution will not be further discussed. The second coordination shell is composed of 3 P atoms at calculated distances of 3.55 Å.

Comparison between *ab initio* MD and fitted EXAFS data brings additional information on the Pu coordination sphere in PEI-MP-Pu. Pu-O and Pu---P distances obtained from the EXAFS data fitting

are less than 0.1 Å different from the values obtained with *ab initio* MD displayed in Table 2 (EXAFS: Pu-O = 2.26(1) Å, Pu---P = 3.62(12) Å; *ab initio* MD: Pu-O_{PEI-MP} = 2.20 Å, Pu-O_{average} = 2.35 Å, Pu---P = 3.55 Å). With *ab initio* MD, an average model of coordination number of 3O_{PEI-MP} + 3O_{wat} + 1N + 3P complexation site (CN = 7+3) was obtained. Note that a coordination number of 7 for Pu(IV) would be at the lower limit of typical coordination numbers for An(IV) (from 8 to 12 depending on the element and its environment).⁶¹ However as PEI-MP* stands for a monomeric unit of PEI-MP terminated with an ethyl and a methyl group, the system is distorted. The corresponding nitrogen atom is further from the actinide ion than it should be. In the real polymer, this N atom would be closer and the coordination would be larger than 7. The EXAFS adjustment also shows a complexation site formed by phosphonate groups with 9 (fixed) O + 2.8(6) P. As mentioned before, the presence (or the absence) of additional water molecules and/or nitrogen atoms in the Pu(IV) first coordination shell is not possible to assess with EXAFS. Since the mixing of O_{wat} and O_{PEI-MP} triggers the average Pu-O distance, a more in-depth comparison between EXAFS and *ab initio* MD is difficult. Nonetheless, EXAFS spectroscopy and *ab initio* MD simulation exhibit Pu-O and Pu---P distances in the same range, confirming the implication of phosphonate groups in the complexation of Pu(IV) in a monodentate pattern.

Another point of comparison comes from literature data although almost no structure is available for organic phosphate Pu complexes. An example is given by Andreev *et al.* who described the structure of a Pu-nucleotide complex (deoxycytidinemonophosphate acid, H₂dCMP).⁶² In this structure, there are 2 independent Pu atoms with similar environments: three O atoms of the monodentate phosphates belonging to three HdCMP⁻ anions, two bidentate nitrate anions, and two water molecules. The three O-phosphate atoms are located at an average of 2.21 Å. Each phosphate group bridges three Pu atoms with an average Pu---P distance equal to 3.62 Å. The structure of Pu(CH₃PO₃)₂ also exhibits a bridging pattern of phosphonate groups that create Pu-O octahedra with average Pu-O distances equal to 2.22 Å. The average Pu---P distance equals 3.53 Å.⁶³ In the same publication the authors also describe the structure of Pu[CH₂(PO₃)₂](H₂O) in which PuO₇ polyhedra are formed.⁶³ The Pu-O bond distances range from 2.18 to 2.30 Å for the Pu-O phosphate bonds and 2.49 Å for the terminal water molecule.⁶² The Pu-O distance proposed in Table 1 (2.26(1) Å) falls in the same (upper) range. The Pu---P distance in PEI-MP-Pu provided in Table 1 (3.62(12) Å) is more difficult to compare because it is impossible to know if the phosphonate groups are bridging or not. Again, the distances described in Table 1 fall within the same range as described in the above structures although all reported distances are scattered between 3.5 and 3.7 Å. One should note in any case that the steric constraint associated with the polymeric backbone of PEI-MP may play a role in extending average distances compared to smaller molecules. An overall schematic picture of the Pu coordination sphere in PEI-MP-Pu is provided in Figure 3b.

DISCUSSION ON THE PEI-MC AND PEI-MP SYSTEMS

The uptake curves of Pu(IV) could not be recorded because of radioprotection limitations in our laboratory that make this type of measurement impossible to perform. But the uptake curves of PEI-MC-Th, PEI-MP-Th and DTPA-Th have been reported in previous publications.^{46,50} The curves are now compared in Figure 5abc with a new and homogenized data treatment. To avoid Th(IV) hydrolysis under these conditions, all uptake curves were recorded in the presence of two molecules of nitrilotriacetic acid (NTA) as a protecting ligand (NTA will also mimic carboxylate metabolites present in serum as citric acid or even bicarbonates). All curves are plotted as a function of the ligand concentration expressed in equivalent monomers. They all exhibit a plateau, *b*, around 100% of complexed Th(IV) attesting for a total displacement of the complexation equilibrium towards complexation at excess of chelating functions. The differences in fitting quality factors between the three curves (PEI-MP-Th : $R^2=0.81$ / $\chi^2_r=0.24$; PEI-MC-Th : $R^2=0.95$ / $\chi^2_r=0.12$; DTPA-Th : $R^2=0.98$ / $\chi^2_r=0.03$) originate from the polydispersity of the polymers as well as from possible loss of Th atoms during the additional step of mineralization needed for the Th titration for PEI-MP-Th and PEI-MC-Th (and not for DTPA-Th).

EC50 values (we recall that *EC50* is the titration point at 50% response of the ligand, therefore the lower the *EC50*, the higher the affinity) reported in Table 3 reflect the chelating affinity towards Th(IV). With DTPA, the *EC50* value (0.13 ± 0.01 mM) is significantly lower than for PEI-MC (1.16 ± 0.15 mM) and PEI-MP (0.61 ± 0.17 mM). If we consider the higher uncertainties related to their *EC50* and the 95% confidence interval determined from the fit, both PEI-MP and PEI-MC polymers seem to exhibit a chemical affinity for Th(IV) of the same order (within a factor of 2). On the other hand, the chemical affinity of DTPA for Th(IV) seems higher than that of PEI-MC, -MP. The affinity constant for both polymers has not been determined, but the affinity constant for DTPA-Th provided in the literature is fairly high ($\log\beta = 26.6$).¹⁹ Another point of comparison is given by the affinity constants of the Th(IV) 1:1 complexes in Th-CO₃²⁻ ($\log\beta_{11} = 11.0$) and Th-HPO₄²⁻ ($\log\beta_{11} = 10.8$).^{64,65}

It is to be noted that the *EC50* values discussed here are a function of the monomeric concentrations within PEI-MC, -MP. First, it is important to remember that each polymer monomer can only bear a maximum (depending on the functionalization rate) of three methylcarboxylate, for PEI-MC, or three methylphosphonate, for PEI-MP. On the contrary, the functionalization degree of the reference compound is higher (5 complexing functions for the DTPA). This inevitably results in lower *EC50* values for PEI-MC-Th and PEI-MP-Th. Second, the functionalization rate, close to 100% for PEI-MP⁵⁰ and around 64% for PEI-MC⁴⁷ could also explain these lower values of *EC50* compared to DTPA (pure monomer).

The slope at the *EC50* point (*n* value of the four logistic parameters fitting equation) is also a

parameter to be considered. The slope accounts for a concentration range necessary to go from the minimal to the maximal response, *i.e.* the higher the slope, the narrower the concentration range. This translates as a cooperative complexation mechanism between the different complexing sites. The n value for PEI-MP-Th (1.3 ± 0.4) is slightly lower than the value for PEI-MC-Th (1.8 ± 0.3) and that of DTPA-Th (2.1 ± 0.2). But they all are in the same range with $n > 1$, suggesting a cooperative mechanism from the functional groups. However, a direct conclusion would be adventurous. For instance, we may note that DTPA-Th has a value larger than 1 suggesting cooperativity between the complexation sites. This result is a little bit surprising for a molecular chelating agent unless polynuclear complexes may form in the experimental conditions. *Ab initio* MD simulations as described in the previous section for PEI-MP-Pu provide additional information on the structure of the coordination site in PEI-MC and PEI-MP.⁴⁶ The radial pair distribution functions $g(r)$ are displayed on Figure 4abcd and average calculated distances are displayed in Table 2. As already noted in the previous section for the EXAFS data, actinidic ionic contraction from Th to Pu is observed on all $g(r)$. A clear shift to lower distances is also observed going from PEI-MC to PEI-MP for the first coordination sphere. But exact contraction values are difficult to give because as stated above the shape of the $g(r)$ is not the same (the coordination spheres are not identical). Although this is not definite proof, this is in agreement with the relative decrease of $EC50$ values from PEI-MC to PEI-MP. Coordination sites obtained with AIMD may also be compared. In PEI-MC, the cation coordination sphere is composed of $3O + 3N$ from the PEI-MC monomer and $1O_{\text{wat}}$ from a water molecule ($CN = 7$), very similar to that of DTPA-M ($5O + 3N$, $CN = 8$).¹⁹ We recall that an average model of $3O_{\text{PEI-MP}} + 3O_{\text{wat}} + 1N + 3P$ complexation site ($CN = 7+3$) has been obtained for PEI-MP-Pu. Note that in the case of Th, there is no N atom in the coordination sphere. According to the simulation, the number of water molecules able to enter in the first coordination sphere of Pu(IV) and Th(IV) is in the order: three when it is complexed to the PEI-MP, one when it is complexed to the PEI-MC, and 0 for the DTPA. This suggests again that DTPA is the most adapted chelate and this is consistent with the lowest $EC50$ value.

Our results confirm that the very high chemical affinity of DTPA for An(IV) is the result of very good structural arrangement around the actinide cation.¹⁹ But considering the specific biodistribution of macromolecular objects, the use of polymeric analogues of molecular chelating agents, such as PEI-MC or PEI-MP, is an interesting strategy to master body distribution and target specific organs. These polymers also have the distinctive ability of being able to complex not only An(IV) (Pu(IV) and Th(IV) in this report) but also actinyls such as $\{UO_2^{2+}\}$ as well.^{47,50} Such approaches could then open the field for a new class of decorporating agents which may complement already existing therapies based on DTPA.

CONCLUSION

Plutonium (Pu) is an anthropogenic element involved in the nuclear industry cycle. Located at the bottom of the periodic table within the actinide family, it is a chemical toxic but also a radiological toxic. This is why Pu decorporation is a major topic of concern in the nuclear industry. In this regard, we have taken a closer look at polymeric chelates and in particular polyethyleneimine (PEI) analogues of DTPA, which may present a real breakthrough in the plutonium decorporation strategy because of their indirect vectorization properties correlated with a specific biodistribution into the lungs, bone, kidney or liver.

In its first section, the present *forum* article describes the complexation site of Th(IV) and Pu(IV) with the PEI-MP polymer using a combination of *ab initio* MD calculations and EXAFS spectroscopy in a pseudo-physiological medium. A similar complexation site for both Th(IV) and Pu(IV) was observed and involves monodentate phosphonates. The presence of O and P atoms in the first and second coordination shells of M = Th(IV), Pu(IV) was determined *via* EXAFS spectroscopy and confirmed by *ab initio* MD calculations. EXAFS best fit distances and *ab initio* MD distances agree within 0.1 Å. Distances in both PEI-MP-Pu and PEI-MP-Th complexes are very similar to the intermolecular distances found in other Pu-phosphonate complexes.^{61, 62}

In the second part of the report, uptake curves of Th(IV) with PEI-MC, PEI-MP and DTPA were also compared. *EC50* values reflect the chelating affinity towards Th(IV) and are ranked in the order DTPA << PEI-MP < PEI-MC. According to the *ab initio* MD simulation, the number of water molecules able to enter in the first coordination sphere of Pu(IV) and Th(IV) when it is complexed to the PEI-MP is equal to three, one for the PEI-MC, and 0 for the DTPA. Pair radial distribution functions *g(r)* show a clear shift to lower distances going from PEI-MC to PEI-MP for the first coordination sphere. Although not a definite proof, this is all in agreement with the relative decrease of *EC50* values from PEI-MC to PEI-MP.

Our results confirm that the very high chemical affinity of DTPA for An(IV) results in a very good fitting around the actinide cation. However, when considering the specific biodistribution of macromolecular objects, the use of polymeric chelating analogues, such as PEI-MC or PEI-MP, could be an interesting strategy to target specific sequestering compartments/organs in the case of actinide contamination. Such physical chemical approaches could then open the field for a new class of decorporating agents, complementing already existing therapies. But of course, a forthcoming development of this approach is to perform *in vitro* toxicity test of PEI-MC and PEI-MP.

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Table 1: EXAFS best fit parameters for the PEI-MP-Tg, PEI-MP-Pu, PEI-MC-Th and PEI-MC-Pu complexes under pseudo biological conditions. Coordination numbers were fixed to 9 for the PEI-MP complex and to 8 for the PEI-MC complex (numbers in italics). Numbers in brackets are the estimated uncertainties. σ^2 is the Debye Waller factor of scattering path. S_0^2 is the global amplitude factor, e_0 is the energy threshold. R_{factor} is the quality factor of the fit in % and Q is the reduced χ^2_r factor of the fit.

Sample	1 st coordination shell	2 nd coordination shell	Fit parameters
PEI-MP-Th (from ref 50)	9 O at 2.35 (1) Å $\sigma^2 = 0.0096 \text{ Å}^2$	2.8(6) P at 3.90 (1) Å $\sigma^2 = 0.0314 \text{ Å}^2$ 1.8 (4) Cl at 3.12 (2) Å $\sigma^2 = 0.0092 \text{ Å}^2$	$S_0^2 = 1.0$ $e_0 = -3.29 \text{ eV}$ $R_{\text{factor}} = 0.5\%$ $Q = 21 \times 10^{-3}$
PEI-MP-Pu	9 O at 2.26 (1) Å $\sigma^2 = 0.0126 \text{ Å}^2$	2.2(9) P at 3.62 (12) Å $\sigma^2 = 0.0149 \text{ Å}^2$ 1.4 (2) Cl at 3.07 (3) Å $\sigma^2 = 0.0055 \text{ Å}^2$	$S_0^2 = 0.9$ $e_0 = 4.89 \text{ eV}$ $R_{\text{factor}} = 2.9\%$ $Q = 4.1$
PEI-MC-Th (from ref 46)	5 O at 2.40 (1) Å $s^2 = 0.0038 \text{ Å}^2$ 3 N at 2.80(2) Å $s^2 = 0.0360 \text{ Å}^2$	5 C $_{\alpha}$ at 3.42 (1) Å $s^2 = 0.0086 \text{ Å}^2$ 9 C $_{\beta}$ at 3.58 (2) Å $s^2 = 0.0086 \text{ Å}^2$ 2.5 (5) Cl at 3.88 (2) Å $s^2 = 0.0091 \text{ Å}^2$	$S_0^2 = 1.0$ $e_0 = 6.26 \text{ eV}$ $R_{\text{factor}} = 1.0\%$ $Q = 0.05$
PEI-MC-Pu (from ref 46)	5 O at 2.34 (1) Å $s^2 = 0.0093 \text{ Å}^2$ 3 N at 2.66 (2) Å $s^2 = 0.0171 \text{ Å}^2$	5 C $_{\alpha}$ at 3.38 (1) Å $s^2 = 0.0116 \text{ Å}^2$ 9 C $_{\beta}$ at 3.45 (2) Å $s^2 = 0.0116 \text{ Å}^2$	$S_0^2 = 1.0$ $e_0 = 5.52 \text{ eV}$ $R_{\text{factor}} = 2.1\%$ $Q = 0.022$

Table 2: Average interatomic distances calculated by *ab initio* MD for the hydrated model of PEI-MP* and PEI-MC* (from ref 46) complexes with Th(IV) and Pu(IV).

Sample	An(IV)-O	An(IV)-N	An(IV)-P
PEI-MP*-Pu	3 O at 2.20 Å ($O_{\text{PEI-MP}}$) 3 O at 2.50 Å (O_{water})	1 N at 2.65 Å	3 P at 3.55 Å
PEI-MP*-Th	3 O at 2.15 Å ($O_{\text{PEI-MP}}$) 3 O at 2.35 Å (O_{water})	-	3 P at 3.45 Å
PEI-MC*-Pu	3 O at 2.22 Å ($O_{\text{PEI-MC}}$) 1 O at 2.28 Å (O_{water})	3 N at 2.68 Å	-
PEI-MC*-Th	3 O at 2.33 Å ($O_{\text{PEI-MC}}$) 1 O at 2.32 Å (O_{water})	3 N at 2.74 Å	-

Table 3: Best fit parameter obtained from the four-parameter logistic equation (eq1) with a is the response at step = 0, b is the maximum response at infinite step, $EC50$ is the titration point at 50% response, n is the slope at the steepest part of the curve. R^2 and Chi_r^2 are the quality factors of the fit.

	a (%)	b (%)	$EC50$ (mM)	n	Fit parameters
PEI-MP-Th	5	97 ± 13	0.61 ± 0.17	1.32 ± 0.35	$R^2 = 0.81$ $\text{Chi}_r^2 = 0.24$
DTPA-Th	5	105 ± 3	0.13 ± 0.007	2.09 ± 0.23	$R^2 = 0.98$ $\text{Chi}_r^2 = 0.03$
PEI-MC-Th	5	110 ± 9	1.16 ± 0.15	1.78 ± 0.26	$R^2 = 0.95$ $\text{Chi}_r^2 = 0.12$

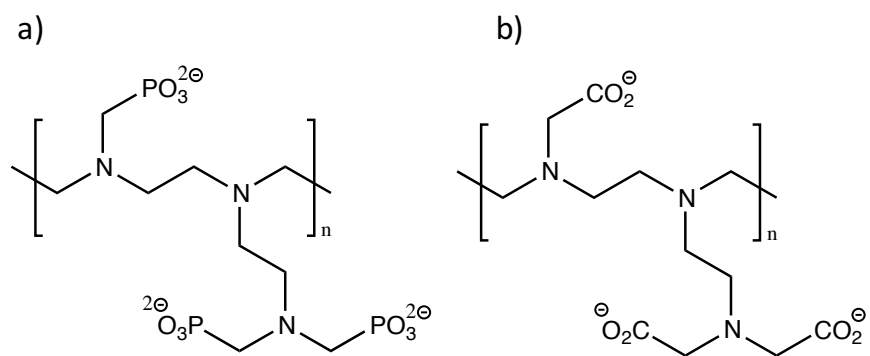


Figure 1ab: Chemical scheme of the monomer of PEI-MP (a) and PEI-MC (b) polymers with an arbitrary charge level.

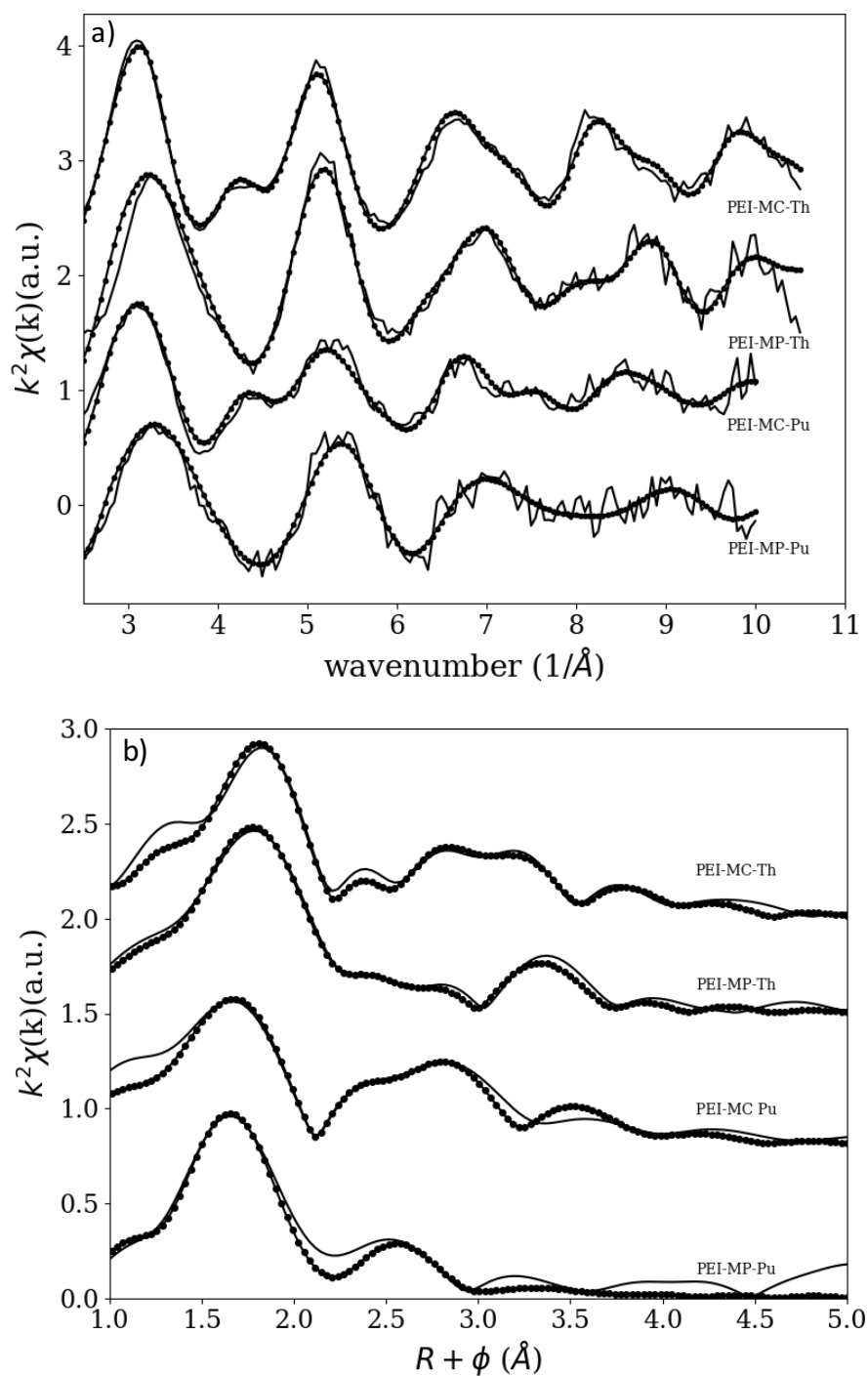


Figure 2ab: EXAFS spectra of the PEI-MP-Pu and PEI-MP-Th complexes under pseudo physiological conditions (a); Corresponding Fourier transform of the EXAFS spectra (b). Solid line = experimental spectrum, dotted line = adjustment. Spectra have been shifted in ordinates for clarity.

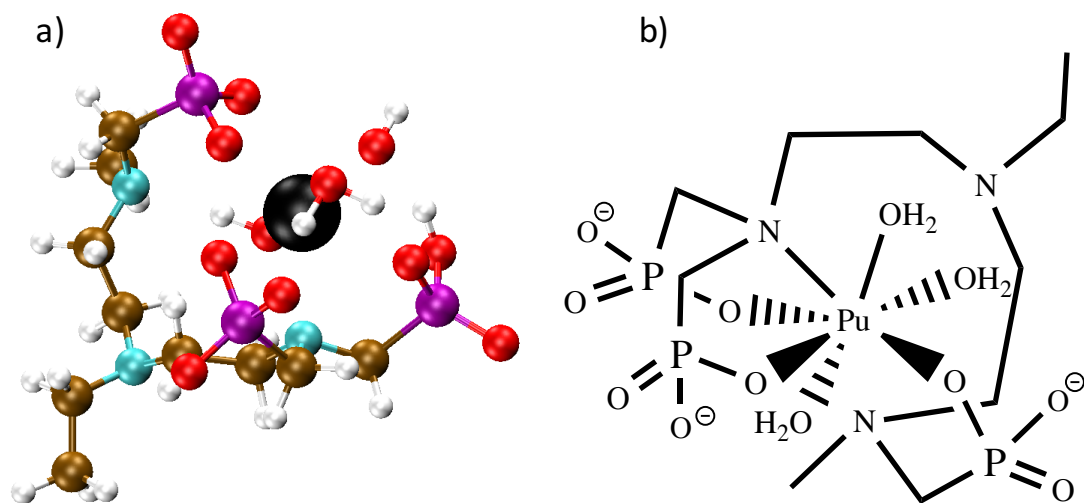


Figure 3ab: Snapshot of PEI-MP*-Pu complex obtained from *ab initio* MD simulation, with plutonium in black, phosphorus in purple, oxygen in red, nitrogen in blue, carbon in brown and hydrogen in white (a); and schematic drawing of the Pu coordination site in PEI-MP-Pu complex (b).

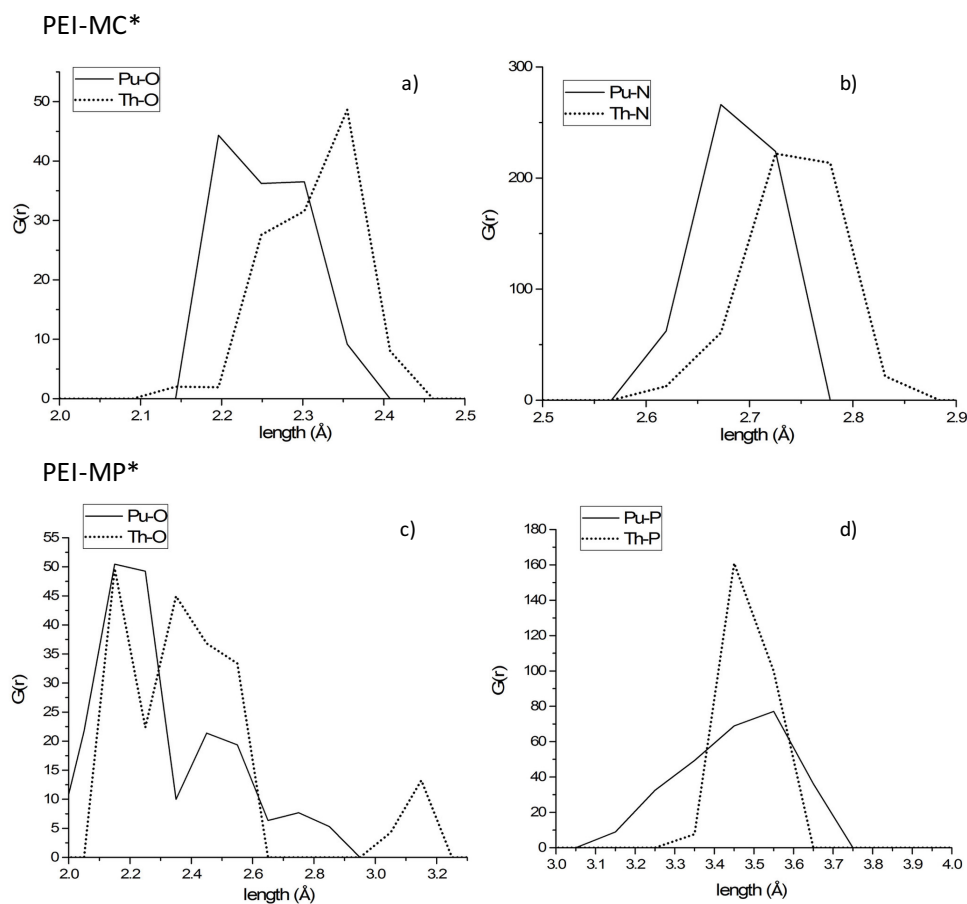
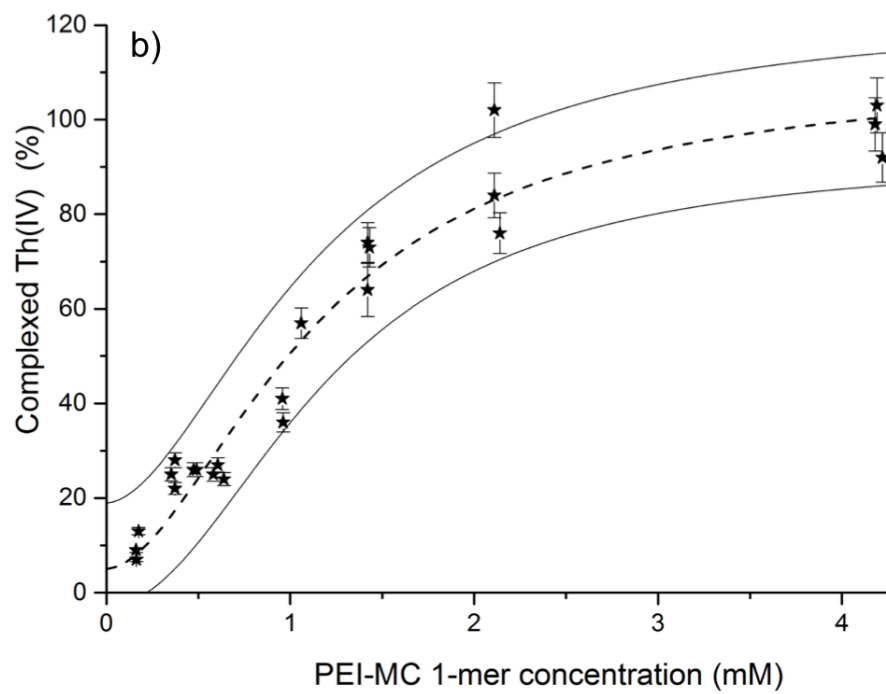
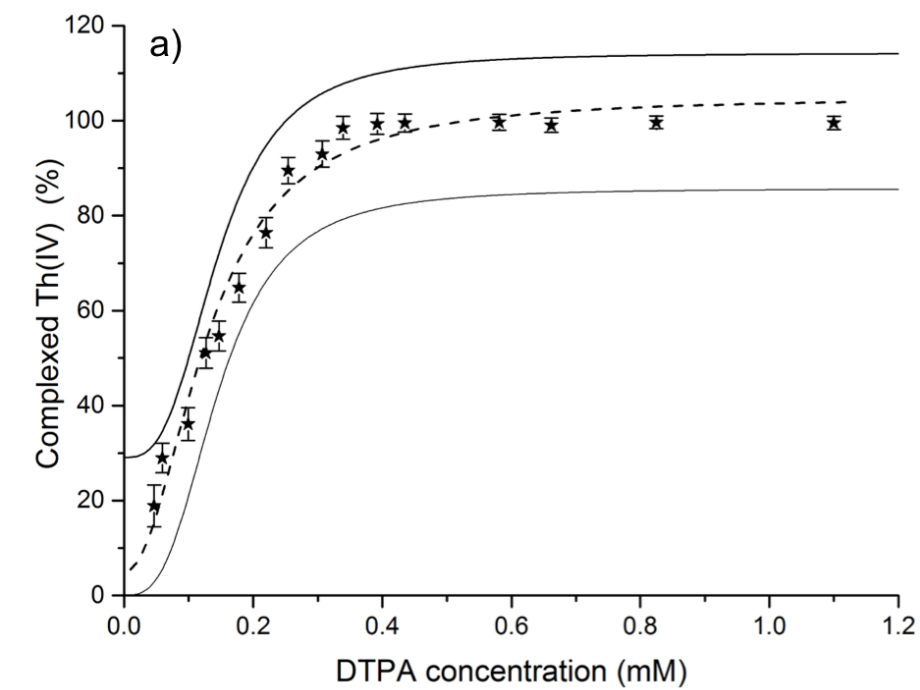


Figure 4abcd: Radial Pair Distribution Functions $g(r)$ of PEI-MC*-M (reproduced from ref. 46 with permission from the Royal Society of Chemistry, a, b) and PEI-MP*-M (c, d) complexes for M = Th(IV), Pu(IV) obtained from *ab initio* MD simulations.



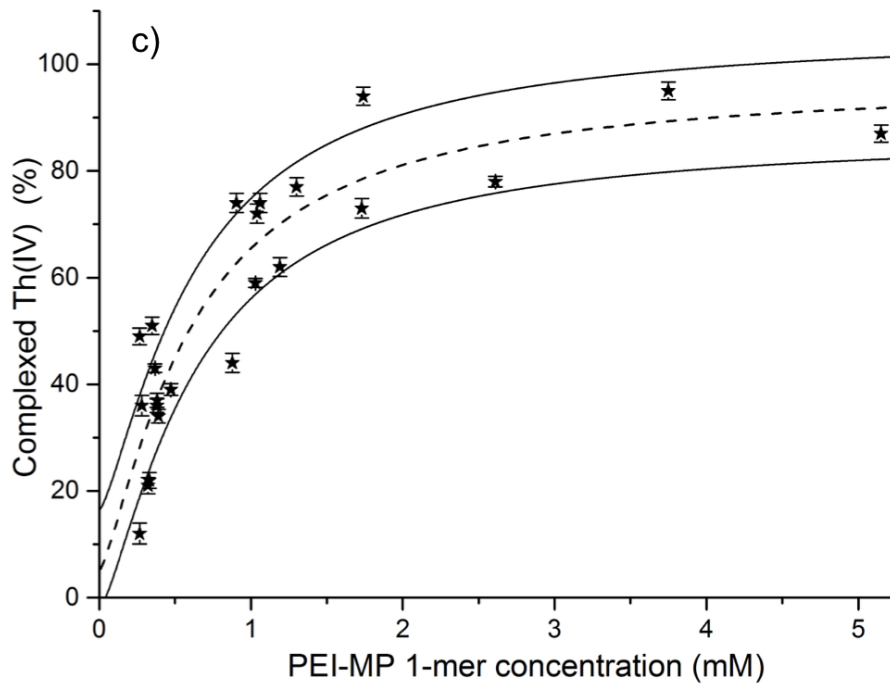


Figure 5abc: Uptake curves representing the complexation rate of Th(IV) as a function of concentration of DTPA (a), PEI-MC (b) and PEI-MP (c) at pH 7.0. Dash lines = fit with the four-parameter logistic non-linear regression model; Black lines = 95% confidence interval.

REFERENCES

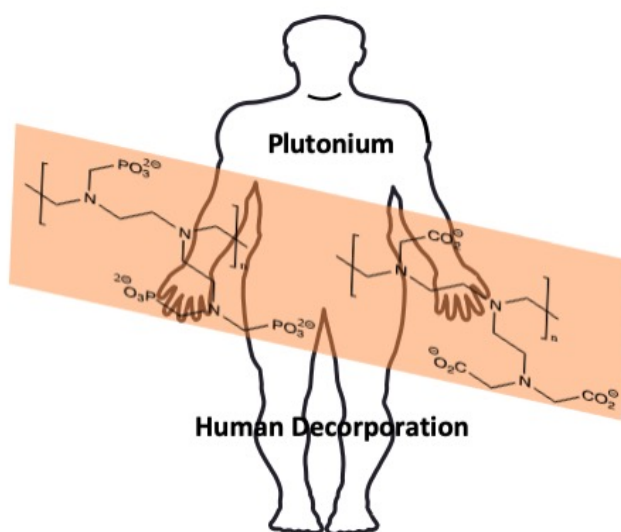
- (1) Schwantes, J. M.; Orton, C. R.; Clark, R. A. Analysis of a nuclear accident: fission and activation product releases from the Fukushima Daiichi nuclear facility as remote indicators of source identification, extent of release, and state of damaged spent nuclear fuel. *Environ. Sci. Technol.* **2012**, *46*, 8621–8627.
- (2) Thakur, P.; Khaing, H.; Salminen-Paatero, S. Plutonium in the atmosphere: A global perspective. *J. Environ. Rad.* **2017**, *39*, 175–176.
- (3) Morrow, P.E.; Witschi, H.; Vore, M.; Hakkinen, P.E.; Mac Gregor, J.; Anders, M.W.; Willhite, C. Profile in toxicology. Harold Carpenter Hodge (1904-1990). *Toxicol. Sci.* **2000**, *53*, 157–158.
- (4) Fattal, E.; Tsapis, F.; Phan, G. Novel drug delivery systems for actinides (uranium and plutonium) decontamination agents, *Adv. Drug Delivery Rev.*, **2015**, *90*, 40–54.
- (5) Leggett, R. W. A Model of the Retention, Translocation and Excretion of Systemic Pu. *Health Phys* **1985**, *49*, 1115–1137.
- (6) Ansoborlo, E.; Prat, O.; Moisy, P.; Auwer, Den, C.; Guilbaud, P.; Carriere, M.; Gouget, B.; Duffield, J.; Doizi, D.; Vercouter, T.; Moulin, C.; Moulin, V. Actinide speciation in relation to biological processes. *Biochimie* **2006**, *88*, 1605–1618.
- (7) Bailey, M. R. The New ICRP Model for the Respiratory Tract. *Radiat Prot Dosimetry* **1994**, *53*, 107–114.
- (8) Wronski, T. J.; Smith, J. M.; Jee, W. S. S. The Microdistribution and Retention of Injected ²³⁹Pu on Trabecular Bone Surfaces of the Beagle: Implications for the Induction of Osteosarcoma. *Radiat. Res.* **1980**, *83*, 74–89.
- (9) Bruenger, F. W.; Stover, B. J.; Stevens, W.; Atherton, D. R. Exchange of ²³⁹Pu IV Between Transferrin and Ferritin in Vitro. *Health Phys.* **1969**, *16*, 339–340.
- (10) Miller, S. C.; Lloyd, R. D.; Bruenger, F. W.; Krahenbuhl, M. P.; Polig, E.; Romanov, S. A. Comparisons of the skeletal locations of putative plutonium-induced osteosarcomas in humans with those in beagle dogs and with naturally occurring tumors in both species. *Radiat. Res.* **2003**, *160*, 517–523.
- (11) Taylor, G. N.; Lloyd, R. D.; Mays, C. W.; Angus, W.; Miller, S. C.; Shabestari, L.; Hahn, F. F. Plutonium- Or Americium-induced Liver Tumors and Lesions in Beagles. *Health Phys.* **1991**, *61*, 337–347.
- (12) Wolfgang, W. G.; Semmler, M.; Möller, W. Dosimetry and Toxicology of Ultrafine Particles, *J. Aerosol Med.* **2004**, *17*, 140–152.
- (13) Schubert, J. Treatment of Plutonium Poisoning by Metal Displacement. *Science*, **1947**, *105*, 389–390.
- (14) Schubert, J. Removal of radioelements from the mammalian body. *Annu. rev. nucl. sci.* **1955**, *5*, 369–412.
- (15) Hursh, J. B. Effect of BAL on Survival of Rats after Lethal Doses of Polonium. *Proc. Soc. Exp. Biol. Med.* **1951**, *79*, 210–212.
- (16) Ansoborlo, R.; Amekraz, B.; Moulin, C.; Moulin, V. R.; Taran, F. D. R.; Bailly, T. O.; Burgada, R.; Heng Napoli, M.-H. L. N.; Jeanson, A. L.; Auwer, Den, C.; Bonin, L.; Moisy, P. Review of actinide decorporation with chelating agents. *C. R. Chim.* **2007**, *10*, 1010–1019.
- (17) Hamilton, J. G.; Scott, K. G. Effect of Calcium Salt of Versene upon Metabolism of Plutonium in the Rat. *Proc. Soc. Exp. Biol. Med.* **2016**, *83*, 301–305.
- (18) Bonin, L.; Aupiais, J.; Kerbaa, M.; Moisy, P.; Topin, S.; Siberchicot, B. Revisiting actinide–DTPA complexes in aqueous solution by CE-ICPMS and ab initio molecular dynamics. *RSC Adv.* **2016**, *6*, 62729–62741.
- (19) Aupiais, J.; Bonin, L.; Auwer, Den, C.; Moisy, P.; Siberchicot, B.; Topin, S. On the use of speciation techniques and ab initio modelling to understand tetravalent actinide behavior in a biological medium: An^{IV} DTPA case. *Dalton Trans.* **2016**, *45*, 3759–3770.
- (20) Durbin, P. W.; Kullgren, B.; Xu, J.; Raymond, K. N. Development of Decorporation Agents for the Actinides. *Radiat. Prot. Dosimetry* **1998**, *79*, 433–443.
- (21) Taylor, G. N.; Lloyd, R. D.; Boseman, J. J.; Atherton, D. R.; Mays, C. W. Removal of

- Plutonium from Beagles Using Ca-DTPA and Zn-DTPA: Effects of Initial DTPA Injection. *Health Phys.* **1978**, *35*, 201–210.
- (22) Stevens, W.; Bruenger, F. W.; Atherton, D. R.; Buster, D. S.; Howerton, G. The Retention and Distribution of ^{241}Am and ^{65}Zn , Given as DTPA Chelates in Rats and of [^{14}C]DTPA in Rats and Beagles. *Radiat. Res.* **1978**, *75*, 397–409.
 - (23) Stather, J. W.; Smith, H.; Bailey, M. R.; Birchall, A.; Bulman, R. A.; Crawley, F. E. The retention of ^{14}C -DTPA in human volunteers after inhalation or intravenous injection. *Health Phys.* **1983**, *44*, 45–52.
 - (24) McAfee, J. G.; Gagne, G.; Atkins, H. L.; Kirchner, P. T.; Reba, R. C.; Blaurock, M. D.; Smith, E. M. Biological distribution and excretion of DTPA labeled with Tc-99m and In-111. *J. Nucl. Med.* **1979**, *20*, 1273–1278.
 - (25) Grémy, O.; Laurent, D.; Coudert, S.; Griffiths, N. M.; Miccoli, L. Decorporation of Pu/Am Actinides by Chelation Therapy: New Arguments in Favor of an Intracellular Component of DTPA Action. *Radiat. Res.* **2016**, *185*, 568–579.
 - (26) Ramounet-Le Gall, B.; Grillon, G.; Rataeu, G.; Burgada, R.; Bailly, T.; Fritsch, P. Comparative decorporation efficacy of 3,4,3-LIHOPO, 4,4,4-LIHOPO and DTPA after contamination of rats with soluble forms of ^{238}Pu and ^{233}U . *Radiat. Prot. Dosimetry* **2003**, *105*, 535–538.
 - (27) Breustedt, B.; Blanchardon, E.; Berard, P.; Fritsch, P.; Giussani, A.; Lopez, M.A.; Luciani, A.; Nosske, D.; Piechowski, J.; Schimmelpfeng, J.; Se randour, A. L. Biokinetic modelling of DTPA decorporation therapy: the conrad approach. *Radiat. Prot. Dosimetry* **2009**, *134*, 38–48.
 - (28) Gorden, A. E. V.; Xu, J.; Raymond, K. N.; Durbin, P. Rational design of sequestering agents for plutonium and other actinides. *Chem. Rev.* **2003**, *103*, 4207–4282.
 - (29) Deblonde, G. J.-P.; Sturzbecher-Hoehne, M.; Abergel, R. J. Solution thermodynamic stability of complexes formed with the octadentate hydroxypyridinonate ligand 3,4,3-LI(1,2-HOPO): a critical feature for efficient chelation of lanthanide(IV) and actinide(IV) ions. *Inorg. Chem.* **2013**, *52*, 8805–8811.
 - (30) Kullgren, B.; Jarvis, E. E.; An, D. D.; Abergel, R. J. Actinide chelation: biodistribution and in vivo complex stability of the targeted metal ions. *Toxicol. Mech. Methods* **2012**, *23*, 18–26.
 - (31) Abergel, R. J.; Durbin, P. W.; Kullgren, B.; Ebbe, S. N.; Xu, J.; Chang, P. Y.; Bunin, D. I. Blakely, E. A.; Bjornstad, K. A.; Rosen, C. J.; Shuh, D. K.; Raymond, K.N. Biomimetic Actinide Chelators: An Update on the Preclinical Development of the Orally Active Hydroxypyridonate Decorporation Agents 3,4,3-LI(1,2-HOPO) and 5-LIO(Me-3,2- HOPO). *Health Phys.* **2010**, *99*, 401–407.
 - (32) Choi, T. A.; Furimsky, A. M.; Swezey, R.; Bunin, D. I.; Byrge, P.; Iyer, L. V.; Chang, P. Y.; Abergel R. J. In Vitro Metabolism and Stability of the Actinide Chelating Agent 3,4,3-LI(1,2-HOPO) *J. Pharm. sci.* **2015**, *104*, 1832–1838.
 - (33) Kelley, M. P.; Deblonde, G. J.-P.; Su, J.; Booth, C. H.; Abergel, R. J.; Batista, E. R.; Yang, P. Bond Covalency and Oxidation State of Actinide Ions Complexed with Therapeutic Chelating Agent 3,4,3-LI(1,2-HOPO). *Inorg. Chem.* **2018**, *57*, 5352–5363.
 - (34) Bulman, R. A.; Griffin, R. J.; Russell, A. T. An examination of some complexing agents for ability to remove intracellularly deposited plutonium. *Health Phys.* **1979**, *37*, 729–734.
 - (35) Miller, S. C.; Bruenger, F. W.; Kuswik-Rabiega, G.; Lloyd, R. D. Decorporation of plutonium by oral administration of a partially lipophilic polyaminocarboxylic acid. *Health Phys.* **1992**, *63*, 195–197.
 - (36) Miller, S. C.; Bruenger, F. W.; Kuswik-Rabiega, G.; Liu, G.; Lloyd, R. D. Duration and dose-related effects of an orally administered, partially lipophilic polyaminocarboxylic acid on the decorporation of plutonium and americium. *J. Pharmacol. Exp. Ther.* **1993**, *267*, 548–554.
 - (37) Sueda, K.; Sadgrove, M. P.; Huckle, J. E.; Leed, M. G. D.; Weber, W. M.; Doyle Eisele, M.; Guilmette, R. A.; Jay, M. Orally Administered DTPA Penta-Ethyl Ester for the Decorporation of Inhaled ^{241}Am . *J. Pharm. Sci.* **2014**, *103*, 1563–1571.
 - (38) Reddy, J. D.; Cobb, R. R.; Dungan, N. W.; Matthews, L. L.; Aiello, K. V.; Ritter, G.; Eppler, B.; Kirk, J. F.; Abernethy, J. A.; Tomisaka, D. M.; Talton, J. D. Preclinical Toxicology, Pharmacology, and Efficacy of a Novel Orally Administered Diethylenetriaminepentaacetic acid (DTPA) Formulation. *Drug Dev. Res.* **2012**, *73*, 232–242.
 - (39) Wilson, J. P.; Cobb, R. R.; Dungan, N. W.; Matthews, L. L.; Eppler, B.; Aiello, K. V.; Curtis,

- S.; Boger, T.; Guilmette, R. A.; Weber, W.; Doyle Eisele, M.; Talton, J. D. Decorporation of Systemically Distributed Americium by a Novel Orally Administered Diethylenetriaminepentaacetic Acid (DTPA) Formulation in Beagle Dogs. *Health Phys.* **2015**, *108*, 308–318.
- (40) Phan, G.; Herbet, A.; Cholet, S.; Benech, H.; Deverre, J.-R.; Fattal, E. Pharmacokinetics of DTPA entrapped in conventional and long-circulating liposomes of different size for plutonium decorporation. *J. Control. Release* **2005**, *110*, 177–188.
 - (41) Phan, G.; Le Gall, B.; Grillon, G.; Rouit, E.; Fouillit, M.; Benech, H.; Fattal, E.; Deverre, J. R. Enhanced decorporation of plutonium by DTPA encapsulated in small PEG-coated liposomes. *Biochimie* **2006**, *88*, 1843–1849.
 - (42) Phan, G.; Le Gall, B.; Deverre, J.-R.; Fattal, E.; Benech, H. Predicting Plutonium Decorporation Efficacy after Intravenous Administration of DTPA Formulations: Study of Pharmacokinetic–Pharmacodynamic Relationships in Rats. *Pharm. Res.* **2006**, *23*, 2030–2035.
 - (43) Zeevaart, J. R.; Louw, W. K. A.; Kolar, Z. I.; Kilian, E.; van Rensburg, F. E. J.; Dormehl, I. C. Biodistribution and Pharmacokinetics of Various Molecular Sized $^{117m}\text{Sn(II)}$ -Polyethyleneiminomethyl Phosphonate Complexes in the Normal Primate Model as Potential Selective Therapeutic Bone Agents. *Arzneimittelforschung* **2011**, *54*, 340–347.
 - (44) Oskuee R. K.; Dehshahri, A.; Shier, W. T.; Ramezani, M. Alkylcarboxylate grafting to polyethylenimine: a simple approach to producing a DNA nanocarrier with low toxicity. *J. Gene. Medicine* **2009**, *11*, 921–932.
 - (45) Wen, S.; Zheng, F.; Shen, M.; Shi, X. Surface modification and PEGylation of branched polyethylenimine for improved biocompatibility. *J. Appl. Polym. Sci.* **2013**, *128*, 3807–3813.
 - (46) Lahrouch, F.; Siberchicot, B.; Leost, L.; Aupiais, J.; Rossberg, A.; Hennig, C.; Auwer, Den, C.; Di Giorgio, C. Polyethylenimine methylenecarboxylate: a macromolecular DTPA analogue to chelate plutonium(IV). *Chem. Commun.* **2018**, *54*, 11705–11708.
 - (47) Lahrouch, F.; Chamayou, A. C.; Creff, G.; Duvail, M.; Hennig, C.; Lozano Rodriguez, M. J.; Auwer, Den, C.; Di Giorgio, C. A Combined Spectroscopic/Molecular Dynamic Study for Investigating a Methyl-Carboxylated PEI as a Potential Uranium Decorporation Agent. *Inorg. Chem.* **2017**, *56*, 1300–1308.
 - (48) Jarvis, N. V.; Zeevaart, J. R.; Wagener, J. M.; Louw, W. K. A.; Dormehl, I. C.; Kilian, E.; Milner, R. J. Metal-ion speciation in blood plasma incorporating the water-soluble polymer, polyethylenimine functionalised with methylenephosphonate groups, in therapeutic radiopharmaceuticals. *Radiochim. Acta* **2002**, *90*, 237–246.
 - (49) Dormehl, I. C.; Louw, W. K. A.; Milner, R. J.; Kilian, E.; Schneeweiss, F. H. A. Biodistribution and Pharmacokinetics of Various Sized Molecular Radiolabelled Polyethyleniminomethyl Phosphonic Acid as a Selective Bone Seeker for Therapy in the Normal Primate Model. *Arzneimittelforschung* **2011**, *51*, 258–263.
 - (50) Lahrouch, F.; Sofronov, O.; Creff, G.; Rossberg, A.; Hennig, C.; Auwer, Den, C.; Di Giorgio, C. Polyethylenimine methylphosphonate: towards the design of a new class of macromolecular actinide chelating agents in the case of human exposition. *Dalton Trans.* **2017**, *46*, 13869–13877.
 - (51) Krauskopf, K. B. Thorium and rare-earth metals as analogs for actinide elements. *Chem. Geol.* **1986**, *55*, 323–335.
 - (52) Leost, L.; Roques, J.; Van der Meeren, A.; Vincent, L.; Sbirrazzuoli, N.; Hennig, C.; Rossberg, A.; Aupiais, J.; Pagnotta, S.; Auwer, Den, C.; Di Giorgio, C. Towards the development of chitosan nanoparticles for plutonium pulmonary decorporation. *Dalton Trans.* **2018**, *47*, 11605–11618.
 - (53) Ravel, B.; Newville, M.; IUCr. ATHENA, ARTEMIS, HEPHAESTUS: data analysis for X-ray absorption spectroscopy using IFEFFIT. *J. Synchrotron Radiat.* **2005**, *12*, 537–541.
 - (54) Rehr, J. J.; Kas, J. J.; Vila, F. D.; Prange, M. P.; Jorissen, K. Parameter-free calculations of X-ray spectra with FEFF9. *Phys. Chem. Chem. Phys.* **2010**, *12*, 5503–5513.
 - (55) Gonze, X.; Amadon, B.; Anglade, P. M.; Beuken, J. M.; Bottin, F.; Boulanger, P.; Bruneval, F.; Caliste, D.; Caracas, R.; Côté, M.; Deutsch, T.; Genovese, L.; Ghosez, P.; Giantomassi, M.; Goedecker, S.; Hamann, D. R.; Hermet, P.; Jollet, F.; Jomard, G.; Leroux, S.; Mancini, M.; Mazevet, S.; Oliveira, M. J. T.; Onida, G.; Pouillon, Y.; Rangel, T.; Rignanese, G. M.; Sangalli,

- D.; Shaltaf, R.; Torrent, M.; Verstraete, M. J.; Zerah, G.; Zwanziger, J. W. ABINIT: First-principles approach to material and nanosystem properties. *Comput. Phys. Commun.* **2009**, *180*, 2582–2615. And pseudo potential reference site : <https://www.abinit.org/psp-tables>
- (56) Jollet, F.; Torrent, M.; Holzwarth, N. Generation of Projector Augmented-Wave atomic data: A 71 element validated table in the XML format. *Comput. Phys. Commun.* **2014**, *185*, 1246–1254.
- (57) Holzwarth, N. A. W.; Tackett, A. R.; Matthews, G. E. A Projector Augmented Wave (PAW) code for electronic structure calculations, Part I: atompaw for generating atom-centered functions. *Comput. Phys. Commun.* **2001**, *135*, 329–347.
- (58) Perdew, J. P.; Burke, K.; Ernzerhof, M. Generalized Gradient Approximation Made Simple. *Phys. Rev. Lett.* **1996**, *77*, 3865–3868.
- (59) Jomard, G.; Amadon, B.; Bottin, F.; Torrent, M. Structural, thermodynamic, and electronic properties of plutonium oxides from first principles. *Phys. Rev. B* **2008**, *78*, 075125-1–075125-11.
- (60) Von Harpe, A.; Petersen, H.; Li, Y.; Kissel, T. Characterization of commercially available and synthesized polyethylenimines for gene delivery. *J. Controlled Release*, **2000**, *69*, 309–322
- (61) Szabó, Z.; Toraishi, T.; Mallet, V.; Grenthe, I. Solution coordination chemistry of actinides: Thermodynamics, structure and reaction mechanisms. *Coord. Chem. Rev.*, **2006**, *250*, 784–815.
- (62) Andreev, G.; Budantseva, N.; Sokolova, M.; Tananaev, I.; Myasoedov, B. Interaction of Transuranium Elements with Biologically Important Ligands: Structural and Spectroscopic Evidence for Nucleotide Coordination to Plutonium. *Inorg. Chem.* **2009**, *48*, 2343–2345.
- (63) Nelson, A.-G. D.; Bray, T. H.; Stanley, F. A.; Albrecht-Schmitt T. E. Periodic Trends in Actinide Phosphonates: Divergence and Convergence between Thorium, Uranium, Neptunium, and Plutonium Systems. *Inorg. Chem.* **2009**, *48*, 4530–4535.
- (64) Santschi, P. H.; Murray, J. W.; Baskaran, M.; Benitez-Nelson, C. R.; Guo, L. D.; Hung, C. C.; Lamborg, C.; Moran, S. B.; Passow, U.; Roy-Barman, M. Thorium speciation in seawater. *Mar. Chem.* **2006**, *100*, 250–268.
- (65) Langmuir, D.; Herman, J. S. The mobility of thorium in natural waters at low temperatures. *Geochim. Cosmochim. Acta* **1980**, *44*, 1753–1766.

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Discussion on alternative polyethylenimine based platforms that may represent a breakthrough in the field of plutonium decorporation field. Combination of *ab initio* Molecular Dynamics and Extended X-ray Absorption Fine Structure to clarify the actinide complexation mechanism.